

5 HUMANIZED ANTIBODIES THAT RECOGNIZE VEROTOXIN II
AND CELL LINE PRODUCING SAME

CROSS REFERENCE TO RELATED APPLICATION

10 This application derives priority from USSN 60/086,570
file May 20, 1998, which is incorporated by reference in its
entirety for all purposes.

FIELD OF INVENTION

The present invention relates generally to the combination of recombinant DNA and monoclonal antibody technologies for developing novel biologics and, more particularly, for example, to the production of non-immunogenic (in humans) immunoglobulins specific for the Verotoxin II (VT2) antigen and Verotoxin II variant antigens (VT2V) and their uses *in vitro* and *in vivo*. The present invention also relates more specifically to humanized monoclonal antibodies against VT2 which are capable of neutralizing VT2 and VT2V, polynucleotide sequences encoding the antibodies, a method of producing the antibodies, pharmaceutical compositions comprising the antibody as an active ingredient, therapeutic agents for treating Verotoxin Producing *E. coli* (VTEC) infection and Hemolytic Uremic Syndrome (HUS) comprising the antibody as an active ingredient, and methods of treating such diseases.

BACKGROUND OF THE INVENTION

Verotoxin (VT), also known as Shiga-like toxin (SLT), is known to cause bloody diarrhea and development of hemolytic uremic syndrome in Verotoxin Producing *E. coli* (VTEC) infection. One of the etiologic agents of VTEC infection is

the virulent *E. coli* 0157. VT may also be produced by bacteria other than those that cause VTEC infection which may also result in a toxic syndrome in humans. In children or elderly adults with reduced immune responses, VT produced by bacteria growing in the intestines may get into the bloodstream by disrupting the intestinal epithelial cells. This may induce Hemolytic Uremic Syndrome (HUS) which is characterized by renal dysfunction and sometimes brain damage (see, e.g., Karmali, M. et al., The Lancet, 1, 619-620 (1983); Siegler, R., The Journal of Pediatrics, 125, 511-518(1994)). To date, there exists no effective drug for these toxic syndromes. Antibiotics have not demonstrated efficacy to prevent the progression of the toxic syndromes (see, e.g., Carter, A. et al., The New England Journal of Medicine, 316, 1496-1500 (1987); Griffen, P. et al., Annals of Internal Medicine, 109, 705-712 (1988)). This might be because of the release of VT from bacteria killed by the antibiotics and the ineffectiveness of antibiotics against VT.

There are two types of Verotoxin, (or Shiga-like toxin) Verotoxin I (VT1 or SLT-1) and Verotoxin II (VT2 or SLT-2). (See, O'Brien et al., Curr. Top. Microbiol. Immunol., 180, 65-94.(1992)). VT2 producing *E. coli* have been isolated from patients suffering from VTEC infection. (See, Russmann et al., J. Med. Microbiol., 40(5), 338-343 (1994)). Ostroff et al., J. Infect. Dis. 160, 994-998 (1989) and Kleanthous et al., Arch. Dis. child. 65, 722-727 (1990) report that *E. coli* 0157 strains that contained VT2 but not VT1 were more frequently associated with HUS. There are additional VT2 variants (VT2V) that have also been isolated clinically. (See, e.g., Microb. Pathog., 8, 47-60 (1990); FEBS Lett., 79, 27-30 (1991); Microb. Pathog., 5, 419-426 (1988); and J. Bacteriol., 170, 4223-4230 (1988)). Armstrong et al., J. Infect. Dis., 171(4), 1042-1045 (1995), have tested a VT absorbent in clinical trials, however, this drug works only in the intestine and is not available to absorb VT that has reached the bloodstream. A γ -globulin preparation showed very low neutralization

activity for VT2 in comparison with that for VT1 (Ashkenazi, S. et al., The Journal of Pediatrics, 113, 1008-1014 (1988); Morooka, T. et al., Acta Paediatrica Japonica, 38, 294-295 (1996)). A mouse monoclonal antibody which neutralizes VT2
5 has been reported. However, this antibody was reported to show a relatively low binding affinity for VT2V (, Schmitt, C. et al., Infection and Immunity, 59, 1065-1073 (1991)).

Further, the use of murine monoclonal antibodies such as those described above have certain drawbacks in human
10 treatment, particularly in repeated therapeutic regimens as explained below. Mouse monoclonal antibodies, for example, tend to have a short half-life in humans, and lack other important immunoglobulin functional characteristics when used in humans.

15 In addition, murine monoclonal antibodies contain substantial amino acid sequences that will be immunogenic when injected into a human patient. Numerous studies have shown that, after injection of a foreign antibody, the immune response elicited by a patient against the injected antibody
20 can be quite strong, essentially eliminating the antibody's therapeutic utility after an initial treatment. Moreover, if mouse or other antigenic (to humans) monoclonal antibodies are used to treat various human diseases, subsequent treatments with unrelated mouse antibodies may be ineffective or even
25 dangerous in themselves, because of cross-reactivity.

While the production of so-called "chimeric antibodies" (e.g., mouse variable regions joined to human constant regions) has proven somewhat successful, a significant immunogenicity problem remains. In general, the production of
30 human immunoglobulins reactive with VT2 antigen with high affinity, as with many antigens, would be extremely difficult using typical human monoclonal antibody production techniques.

Thus, there is a need for improved forms of humanized immunoglobulins specific for VT2 antigen that are
35 substantially non-immunogenic in humans, yet easily and economically produced in a manner suitable for therapeutic

formulation and other uses. The present invention fulfills these and other needs.

SUMMARY OF THE INVENTION

5 The invention provides humanized antibodies that specifically bind to VT2 and/or VT2 variant. Some such antibodies specifically bind to the B subunit of VT2 and/or the B subunit of VT2 variant. Some such antibodies neutralizes VT2 and/or VT2 variant. Preferred antibodies
10 neutralized VT2 and/or VT2 variants such as to provide at least 50%, 75%, 95% or 100% protection against mice or other mammalian subject challenged with 10 LD₅₀ of VT2 or VT2 variant.

15 Some humanized antibodies are a humanized form of mouse antibody VTm1-1, the mouse antibody being characterized by a light chain variable region shown in Fig. 1B and a heavy chain variable region shown in Fig. 1A.

20 The invention further provides antibodies that competes with mouse antibody VTm1-1 for specific binding to VT2 and/or VT2 variant.

Some of the humanized antibodies, as described above, comprise complementarity determining regions from the mouse VTm1-1 antibody and heavy and light chain variable region frameworks from the human GF4 antibody heavy and light chain
25 frameworks, provided that at least one position selected from the group consisting of L49, H29, H30, H49 and H98, is occupied by the amino acid present in the equivalent position of the mouse VTm1-1 antibody heavy or light chain variable region framework. Such humanized antibodies specifically
30 binds to verotoxin II with an affinity constant between 10^{-7} M⁻¹ and three, five or ten-fold the affinity of the mouse VTm1-1 antibody.

35 In some humanized antibodies described in the previous paragraph, each position selected from the group consisting of L49, H29, H30, H49 and H98 is occupied by the amino acid

present in the equivalent position of the mouse VTm1-1 antibody heavy or light chain variable region framework.

In some of the above humanized antibodies, at least one position selected from the group L3, L4, L19, L76, L79, L85, H1, H4, H5, H79, H89 and H93 is occupied by an amino acid present in the equivalent position of a human antibody heavy or light chain consensus sequence. In some humanized antibodies each position selected from the group L3, L4, L19, L76, L79, L85, H1, H4, H5, H79, H89 and H93 is occupied by an amino acid present in the equivalent position of a human antibody heavy or light chain consensus sequence.

Some humanized antibodies comprise a heavy chain variable region shown in Fig. 2A and a light chain variable region shown in Fig. 2B provided that one or more positions selected from the group consisting of L49, H29, H30, H49, H98, L3, L4, L19, L76, L79, L85, H1, H4, H5, H79, H89, and H93 may be substituted as shown in Tables 2 and 3.

Some humanized antibodies comprise a heavy chain variable region shown in Fig. 2A and a light chain variable region shown in Fig. 2B.

Some humanized antibodies comprise a humanized heavy chain having at least 85% identity with the humanized heavy chain shown in Fig. 2A and a humanized light chain having at least 85% sequence identity with the humanized light chain showing in Fig. 2B, provided that at least one position selected from the group consisting of L49, H29, H30, H49 and H98, is occupied by the amino acid present in the equivalent position of the mouse VTm1-1 antibody heavy or light chain variable region framework.

Some of the humanized antibodies as described above comprise two pairs of light/heavy chain dimers, wherein each chain comprises a variable region and a constant region.

Some of the humanized antibodies as described above are a Fab fragment or a $F(ab')_2$.

Optionally, humanized antibodies, as described above, are provided in purified form.

Some humanized antibodies, as described above, have an IgG₁ immunoglobulin isotype.

The invention further provides methods of producing humanized VTm1-1 antibody. Such methods comprise culturing a cell line, which encodes heavy and light chain chains of any of the antibodies described above, whereby the humanized antibody is expressed; and recovering the humanized antibody expressed by the cell line. Some such methods further comprise mixing the antibody with a pharmaceutically acceptable carrier to produce a pharmaceutical composition.

The invention further provides pharmaceutical compositions comprising any of the antibodies described above and a pharmaceutically acceptable carrier. A preferred such composition comprises a humanized antibody comprising a heavy chain variable region shown in Fig. 2A and a light chain variable region shown in Fig. 2B. The invention further provides for the use of any of the antibodies described above in the manufacture of a medicament for the treatment of a patient suffering or at risk of toxic effects from a verotoxin.

The invention further provides methods of treating a patient suffering or at risk of toxic effects from a verotoxin, comprising administering to the patient an effective dosage of a human or humanized antibody that specifically binds to verotoxin II and/or verotoxin II variant. In some such methods, the antibody competes with mouse antibody VTm1-1 for specific binding to verotoxin II or verotoxin II variant. In some such methods, the humanized antibody specifically binds to VT2 and/or VT2 variant. In some such methods, the humanized antibody specifically binds to the B subunit of VT2 and/or VT2 variant. In some such methods, the humanized antibody specifically binds to VT2 and/or VT2 variant and neutralizes VT2 and/or VT2 variant. In some such methods, the humanized antibody specifically binds to the B subunit of VT2 and/or the B subunit of VT2 variant and neutralizes VT2 and/or VT2 variant. In some such methods,

the antibody is a humanized antibody, which is a humanized form of the mouse VTm1-1 antibody. In some such methods, the antibody is a humanized antibody comprising a heavy chain variable region shown in Fig. 2A and a light chain variable region shown in Fig. 2B. In some such methods, the patient is infected with verotoxin producing *E. coli* and the antibody is administered therapeutically. In some such methods, the patient is at risk of infection by verotoxin producing *E. coli* and the antibody is administered prophylactically. Some such methods further comprise monitoring the patient for recovery from the toxic effects of verotoxin II or verotoxin II variant.

In a further aspect, the invention provides a cell lines which produces any of the above described antibodies.

The present invention provides novel compositions useful, for example, in the treatment of Verotoxin producing *E. coli* (VTEC) infection and Hemolytic Uremic Syndrome (HUS), the compositions containing humanized immunoglobulins specifically capable of binding to the B subunit of VT2 antigen and of neutralizing VT2 and VT2 variants. The immunoglobulins can have two pairs of light chain/heavy chain complexes, at least one chain comprising one or more mouse complementarity determining regions functionally joined to human framework region segments. For example, mouse complementarity determining regions, with or without additional naturally-associated mouse amino acid residues, can be introduced into human framework regions to produce humanized immunoglobulins capable of binding to the antigen at affinity levels stronger than about 10^7 M^{-1} . These humanized immunoglobulins will also be capable of blocking the binding of the CDR-donating mouse monoclonal antibody to VT2.

The immunoglobulins, including binding fragments and other derivatives thereof, of the present invention may be produced readily by a variety of recombinant DNA techniques, with ultimate expression in transfected cells, preferably immortalized eukaryotic cells, such as myeloma or hybridoma

cells. Polynucleotides comprising a first sequence coding for humanized immunoglobulin framework regions and a second sequence set coding for the desired immunoglobulin complementarity determining regions can be produced synthetically or by combining appropriate cDNA and genomic DNA segments.

The humanized immunoglobulins may be utilized in substantially pure form in treating potentially toxic outcomes from VT2 or VT2V such as those produced during Verotoxin producing *E. coli* (VTEC) infection and Hemolytic Uremic Syndrome (HUS). The humanized immunoglobulins or their complexes can be prepared in a pharmaceutically accepted dosage form, which will vary depending on the mode of administration.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1. The cDNA and translated amino acid sequences of the heavy chain (A) and light chain (B) variable regions of the mouse VTm1.1 antibody (MuVTm1.1). The complementarity determining regions (CDRs) are underlined and the first amino acids of the mature chains are double underlined.

Figure 2. The cDNA and translated amino acid sequences of the heavy chain (A) and light chain (B) variable regions of the humanized VTm1.1 antibody (HuVTm1.1). The complementarity determining regions (CDRs) are underlined and the first amino acids of the mature chains are double underlined.

Figure 3. Scheme for synthesis of humanized antibody variable region cDNA.

Figure 4. Competitive binding of MuVTm1.1 and HuVTm1.1 antibodies to *E. coli* verotoxin II (VT2). Increasing concentrations of competitor antibody were incubated with coated VT2 in the presence of biotinylated tracer MuVTm1.1.

Absorbance was measured and plotted against the concentration of the unlabeled competitor antibodies.

Figure 5. *In vitro* neutralizing activity of HuVTm1.1 versus MuVTm1.1.

Figure 6. Identification of Recognized antigen (the B subunit of VT2) of HuVTm1.1.

Figure 7. Neutralizing activity of MuVTm1.1 against VT2 and VT2 variant.

DEFINITIONS

The phrase "substantially identical," in the context of two nucleic acids or polypeptides (e.g., DNAs encoding a humanized immunoglobulin or the amino acid sequence of the humanized immunoglobulin) refers to two or more sequences or subsequences that have at least about 80%, most preferably 85%, 90-95% or higher nucleotide or amino acid residue identity, when compared and aligned for maximum correspondence, as measured using the following sequence comparison method and/or by visual inspection. Such "substantially identical" sequences are typically considered to be homologous. Preferably, the "substantial identity" exists over a region of the sequence that is at least about 50 residues in length, more preferably over a region of at least about 100 residues, and most preferably the sequences are substantially identical over at least about 150 residues, or over the full length of the two sequences to be compared. As described below, any two antibody sequences can only be aligned in one way, by using the numbering system of Kabat. Therefore, for antibodies, percent identity has a unique and well-defined meaning.

Amino acid sequences from the variable regions of the mature heavy and light chains of immunoglobulins are designated Hx and Lx respectively, where x is a number

designating the position of an amino acid according to the scheme of Kabat, Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda MD, 1987 and 1991). Kabat lists many amino acid sequences for antibodies for each subgroup, and lists the most commonly occurring amino acid for each residue position in that subgroup to generate a consensus sequence. Kabat uses a method for assigning a residue number to each amino acid in a listed sequence, and this method for assigning residue numbers has become standard in the field. Kabat's scheme is extendible to other antibodies not included in his compendium by aligning the antibody in question with one of the consensus sequences in Kabat by reference to conserved amino acids. For example, an amino acid at the L50 position of a human antibody occupies the equivalent position to an amino acid at position L50 of a mouse antibody.

Conservative amino acid substitutions refer to the interchangeability of residues having similar side chains. For example, a group of amino acids having aliphatic side chains is glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains is serine and threonine; a group of amino acids having amide-containing side chains is asparagine and glutamine; a group of amino acids having aromatic side chains is phenylalanine, tyrosine, and tryptophan; a group of amino acids having basic side chains is lysine, arginine, and histidine; and a group of amino acids having sulfur-containing side chains is cysteine and methionine. Preferred conservative amino acids substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, and asparagine-glutamine.

Analogs of exemplified antibody sequences can be screened for retention of binding activity using phage display methods as described by, for example, Cesareni, *FEBS Lett* 307:66-70 (1992); Swimmer et al., *Proc. Natl. Acad. Sci. USA* 89:3756-60 (1992); Gram et al., *Proc. Natl. Acad. Sci. USA* 89:3576-80

(1992); Clackson et al., Nature 352:624-8 (1991); Scott & Smith, Science 249:386-90 (1990); Garrard et al., Bio/Techniques 9:1373-1377 (1991), which are incorporated herein by reference in their entirety for all purposes.

5 The term "substantially pure" means an object species is the predominant species present (i.e., on a molar basis it is more abundant than any other individual species in the composition), and preferably a substantially purified fraction is a composition wherein the object species comprises at least
10 about 50 percent (on a molar basis) of all macromolecular species present. Generally, a substantially pure composition will comprise more than about 80 to 90 percent of all macromolecular species present in the composition. Most preferably, the object species is purified to essential
15 homogeneity (contaminant species cannot be detected in the composition by conventional detection methods) wherein the composition consists essentially of a single macromolecular species.

 Competition between antibodies is determined by an assay
20 in which the immunoglobulin under test inhibits specific binding of a reference antibody to an antigenic determinant. Numerous types of competitive binding assays are known, for example: solid phase direct or indirect radioimmunoassay (RIA), solid phase direct or indirect enzyme immunoassay
25 (EIA), sandwich competition assay (see Stahli et al., Methods in Enzymology 9:242-253 (1983)); solid phase direct biotin-avidin EIA (see Kirkland et al., J. Immunol. 137:3614-3619 (1986)); solid phase direct labeled assay, solid phase direct labeled sandwich assay (see Harlow and Lane, "Antibodies, A
30 Laboratory Manual," Cold Spring Harbor Press (1988)); solid phase direct label RIA using I-125 label (see Morel et al., Molec. Immunol. 25(1):7-15 (1988)); solid phase direct biotin-avidin EIA (Cheung et al., Virology 176:546-552 (1990)); and direct labeled RIA (Moldenhauer et al., Scand. J. Immunol.
35 32:77-82 (1990)). Usually the test immunoglobulin is present in excess. Antibodies identified by competition assay

(competing antibodies) include antibodies binding to the same epitope as the reference antibody and antibodies binding to an adjacent epitope sufficiently proximal to the epitope bound by the reference antibody for steric hindrance to occur.

5 Usually, when a competing antibody is present in excess, it will inhibit specific binding of a reference antibody to a designated antigenic target by at least 50 or 75%.

DETAILED DESCRIPTION OF THE INVENTION

10 In accordance with the present invention, humanized immunoglobulins specifically reactive with the B subunit of VT2 are provided. These immunoglobulins, which have binding affinities to the B subunit of VT2 of at least about 10^7 M^{-1} to 10^{10} M^{-1} , and preferably 10^8 M^{-1} to 10^{10} M^{-1} or stronger, are
15 capable of, e.g., neutralizing the toxicity of VT2 and VT2V (the VT2 antigens). The humanized immunoglobulins will have a human framework and will have one or more complementarity determining regions (CDR's) from an immunoglobulin, typically a mouse immunoglobulin, specifically reactive with VT2
20 antigens. In a preferred embodiment, one or more of the CDR's will come from the MuVTm1.1 antibody. Thus, the immunoglobulins of the present invention, which can be produced economically in large quantities, find use, for example, in the treatment of the toxic outcomes from Verotoxin
25 producing *E. coli* (VTEC) infection and Hemolytic Uremic Syndrome (HUS) in human patients by a variety of techniques.

The basic antibody structural unit is known to comprise a tetramer. Each tetramer is composed of two identical pairs of polypeptide chains, each pair having one "light" (about 25kD)
30 and one "heavy" chain (about 50-70kD). The amino-terminal portion of each chain includes a variable region of about 100 to 110 or more amino acids primarily responsible for antigen recognition. The carboxy-terminal portion of each chain defines a constant region primarily responsible for effector
35 function.

Light chains are classified as either kappa or lambda. Heavy chains are classified as gamma, mu, alpha, delta, or epsilon, and define the antibody's isotype as IgG, IgM, IgA, IgD and IgE, respectively. Within light and heavy chains, the variable and constant regions are joined by a "J" region and about 12 or more amino acids, with the heavy chain also including a "D" region or about 10 more amino acids. (See, generally, Fundamental Immunology, Paul, W., Ed., Chapter 7, pgs. 131-166, Raven Press, N.Y. (1984), which is incorporated herein by reference.)

The variable regions of each light/heavy chain pair form the antibody binding site. The chains all exhibit the same general structure of relatively conserved framework regions joined by three hypervariable regions, also called Complementarity Determining Regions or CDR's (see, "Sequences of Proteins of Immunological Interest," Kabat, E., et al., U.S. Department of Health and Human Services, (1987); and Chothia and Lesk, J. Mol. Biol., 196, 901-917 (1987), which are incorporated herein by reference). The CDR's from the two chains of each pair are aligned by the framework regions, enabling binding to a specific epitope.

As used herein, the term "immunoglobulin" refers to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. The recognized immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon and mu constant region genes, as well as the myriad immunoglobulin variable region genes. The immunoglobulins may exist in a variety of forms besides antibodies; including, for example, Fv, Fab, and F(ab')₂ as well as bifunctional hybrid antibodies (e.g., Lanzavecchia et al., Eur. J. Immunol. 17, 105 (1987)) and in single chains (e.g., Huston et al., Proc. Natl. Acad. Sci. U.S.A., 85, 5879-5883 (1988) and Bird et al., Science 242, 423-426 (1988), which are incorporated herein by reference). (See, generally, Hood et al., Immunology, Benjamin, N.Y., 2nd ed. (1984), Harlow and Lane, Antibodies. A Laboratory Manual, Cold Spring Harbor

Laboratory (1988) and Hunkapiller and Hood, Nature, 323, 15-16 (1986), which are incorporated herein by reference.)).

Chimeric antibodies are antibodies whose light and heavy chain genes have been constructed, typically by genetic engineering, from immunoglobulin gene segments belonging to different species. For example, the variable (V) segments of the genes from a mouse monoclonal antibody may be joined to human constant (C) segments, such as γ_1 and γ_3 . A typical therapeutic chimeric antibody is thus a hybrid protein consisting of the V or antigen-binding domain from a mouse antibody and the C or effector domain from a human antibody, although other mammalian species may be used.

As used herein, the term "framework region" refers to those portions of immunoglobulin light and heavy chain variable regions that are relatively conserved (i.e., other than the CDR's) among different immunoglobulins in a single species, as defined by Kabat, et al., op. cit. As used herein, a "human framework region" is a framework region that is substantially identical (about 85% or more) to the framework region of a naturally occurring human antibody.

As used herein, the term "humanized immunoglobulin" refers to an immunoglobulin comprising a human framework, at least one CDR from a non-human antibody, and in which any constant region present is substantially identical to a human immunoglobulin constant region, i.e., at least about 85-90%, preferably at least 95% identical. Hence, all parts of a humanized immunoglobulin, except possibly the CDR's, are substantially identical to corresponding parts of one or more native human immunoglobulin sequences. For example, a humanized immunoglobulin would not encompass a chimeric mouse variable region/human constant region antibody.

Humanized antibodies have at least three potential advantages over mouse and in some cases chimeric antibodies for use in human therapy:

Because the effector portion is human, it may interact better with the other parts of the human immune system

(e.g., destroy the target cells more efficiently by complement-dependent cytotoxicity (CDC) or antibody-dependent cellular cytotoxicity (ADCC)).

The human immune system should not recognize the framework or C region of the humanized antibody as foreign, and therefore the antibody response against such an injected antibody should be less than against a totally foreign mouse antibody or a partially foreign chimeric antibody.

Injected mouse antibodies have been reported to have a half-life in the human circulation much shorter than the half-life of normal antibodies (Shaw, D. et al., J. Immunol., 138, 4534-4538 (1987)). Injected humanized antibodies will presumably have a half-life essentially identical to naturally occurring human antibodies, allowing smaller and less frequent doses to be given.

In one aspect, the present invention is directed to recombinant polynucleotides encoding the heavy and/or light chain CDR's from an immunoglobulin capable of binding the B subunit of VT2, such as monoclonal antibody MuVTm1.1. The polynucleotides encoding these regions will typically be joined to polynucleotides encoding appropriate human framework regions. As to the human framework region, a framework or variable region amino acid sequence of a CDR-providing non-human immunoglobulin is compared with corresponding sequences in a human immunoglobulin sequence collection, and a sequence having high homology is selected. Exemplary polynucleotides, which on expression code for the polypeptide chains comprising the heavy and light chain CDR's of monoclonal antibody MuVTm1.1 are included in Fig. 1. Due to codon degeneracy and non-critical amino-acid substitutions, other polynucleotide sequences can be readily substituted for those sequences, as detailed below. The design of humanized immunoglobulins may be carried out as follows. When an amino acid falls under the

following category, the framework amino acid of a human immunoglobulin to be used (acceptor immunoglobulin) is replaced by a framework amino acid from a CDR-providing non-human immunoglobulin (donor immunoglobulin):

5 the amino acid in the human framework region of the
acceptor immunoglobulin is unusual for human
immunoglobulin at that position, whereas the
corresponding amino acid in the donor immunoglobulin
is typical for human immunoglobulin in that position;
10 the position of the amino acid is immediately adjacent to
one of the CDR's; or
the amino acid is within about 3Å of a CDR in a tertiary
structure immunoglobulin model (see, Queen et al., op.
cit., and Co et al., Proc. Natl. Acad. Sci. USA 88,
15 2869 (1991), respectively, both of which are
incorporated herein by reference).

When each of the amino acids in the human framework
region of the acceptor immunoglobulin and a
corresponding amino acid in the donor immunoglobulin
20 is unusual for human immunoglobulin at that position,
such an amino acid is replaced by an amino acid
typical for human immunoglobulin at that position.

For a detailed description of the production of humanized
25 immunoglobulins see, Queen et al., op. cit., and Co et al.,
op. cit.

The polynucleotides will typically further include an
expression control polynucleotide sequence operably linked to
the humanized immunoglobulin coding sequences, including
30 naturally-associated or heterologous promoter regions.
Preferably, the expression control sequences will be
eukaryotic promoter systems in vectors capable of transforming
or transfecting eukaryotic host cells, but control sequences
for prokaryotic hosts may also be used. Once the vector has
35 been incorporated into the appropriate host, the host is
maintained under conditions suitable for high level expression

of the nucleotide sequences, and, as desired, the collection and purification of the light chains, heavy chains, light/heavy chain dimers or intact antibodies, binding fragments or other immunoglobulin forms may follow.

5 The nucleic acid sequences of the present invention capable of ultimately expressing the desired humanized antibodies can be formed from a variety of different polynucleotides (genomic or cDNA, RNA, synthetic oligonucleotides, etc.) and components (e.g., V, J, D, and C
10 regions), as well as by a variety of different techniques. Joining appropriate genomic and synthetic sequences is presently the most common method of production, but cDNA sequences may also be utilized (see, European Patent Publication No. 0239400 and Riechmann, L. et al., Nature,
15 332, 323-327 (1988), both of which are incorporated herein by reference.)

Human constant region DNA sequences can be isolated in accordance with well known procedures from a variety of human cells, but preferably immortalized B-cells (see, Kabat op. cit.
20 cit. and WP 87/02671). The CDR's for producing the immunoglobulins of the present invention will be similarly derived from monoclonal antibodies capable of binding to VT2 antigens and produced in any convenient mammalian source, including, mice, rats, rabbits, or other vertebrate capable of
25 producing antibodies by well known methods. Suitable source cells for the polynucleotide sequences and host cells for immunoglobulin expression and secretion can be obtained from a number of sources, such as the American Type Culture Collection (Catalogue of Cell Lines and Hybridomas, Fifth
30 edition (1985) Rockville, Maryland, U.S.A., which is incorporated herein by reference).

In addition to the humanized immunoglobulins specifically described herein, other "substantially homologous" modified immunoglobulins can be readily designed and manufactured
35 utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the framework regions

can vary from the native sequences at the primary structure level by several amino acid substitutions, terminal and intermediate additions and deletions, and the like. Moreover, a variety of different human framework regions may be used singly or in combination as a basis for the humanized immunoglobulins of the present invention. In general, modifications of the genes may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Gillman and Smith, Gene 8, 81-97 (1979) and Roberts S. et al., Nature 328, 731-734 (1987), both of which are incorporated herein by reference.)

Alternatively, polypeptide fragments comprising only a portion of the primary antibody structure may be produced, which fragments possess one or more immunoglobulin activities (e.g., complement fixation activity). These polypeptide fragments may be produced by proteolytic cleavage of intact antibodies by methods well known in the art, or by inserting stop codons at the desired locations in the vectors using site-directed mutagenesis, such as after CH1 to produce Fab fragments or after the hinge region to produce F(ab')₂ fragments. Single chain antibodies may be produced by joining VL and VH with a DNA linker (see Huston et al., op cit., and Bird et al., op cit.). Also because like many genes, the immunoglobulin-related genes contain separate functional regions, each having one or more distinct biological activities, the genes may be fused to functional regions from other genes to produce fusion proteins having novel properties.

As stated previously, the polynucleotides will be expressed in hosts after the sequences have been operably linked to (i.e., positioned to ensure the functioning of) an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly, expression vectors will contain selection marker, e.g., tetracycline or neomycin, to permit detection of those cells

transformed with the desired DNA sequences (see, e.g., U.S. Patent 4,704, 362, which is incorporated herein by reference).

E. coli is one prokaryotic host useful particularly for cloning the polynucleotides of the present invention. Other microbial hosts suitable for use include bacilli, such as Bacillus subtilus, and other enterobacteriaceae, such as Salmonella, Serratia, and various Pseudomonas species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (trp) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing transcription and translation.

Other microbes, such as yeast, may also be used for expression. Saccharomyces is a preferred host, with suitable vectors having expression control sequences, such as promoters, including 3-phosphoglycerate kinase or other glycolytic enzymes, and an origin of replication, termination sequences and the like as desired.

In addition to microorganisms, mammalian tissue cell culture may also be used to express and produce the polypeptides of the present invention (see, Winnacker, From Genes to Clones, VCH Publishers, N.Y., N.Y. (1987), which is incorporated herein by reference). Eukaryotic cells are actually preferred, because a number of suitable host cell lines capable of secreting intact immunoglobulins have been developed in the art, and include the CHO cell lines, various COS cell lines, HeLa cells, preferably myeloma cell lines, etc., or transformed B-cells of hybridomas. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter, and

enhancer (Queen et al., Immunol. Rev. 89, 46-68 (1986), which is incorporated herein by reference), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences are promoters derived from immunoglobulin genes, SV40, Adenovirus, Bovine Papilloma Virus, cytomegalovirus and the like.

The vectors containing the polynucleotide sequences of interest (e.g., the heavy and light chain encoding sequences and expression control sequences) can be transferred into the host cell by well-known methods, which vary depending on the type of cellular host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cellular hosts. (See, generally, Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press (1982), which is incorporated herein by reference.)

Once expressed, the whole antibodies, their dimers, individual light and heavy chains, or other immunoglobulin forms of the present invention can be purified according to standard procedures in the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like (see, generally, Scopes, R., Protein Purification, Springer-Verlag, N.Y. (1982), which is incorporated herein by reference). Substantially pure immunoglobulins of at least about 90 to 95% homogeneity are preferred, and 98 to 99% or more homogeneity most preferred, for pharmaceutical uses. Once purified, partially or to homogeneity as desired, the polypeptides may then be used therapeutically (including extracorporeally) or in developing and performing assay procedures, immunofluorescent stainings, and the like. (See, generally, Immunological Methods, Vols. I and II, Lefkovits and Pernis, eds., Academic Press, New York, N.Y. (1979 and 1981).

The immunoglobulins of the present invention will typically find use individually in treating the toxic effects of Verotoxinproducing *E. coli* (VTEC) infection and Hemolytic Uremic Syndrome (HUS) and/or in neutralizing VT2 antigens. By way of example but not limitation, some typical disease states
5 suitable for treatment include hemorrhagic colitis locally in the gut, renal dysfunction, and brain damage.

The humanized immunoglobulins and pharmaceutical compositions thereof of this invention are particularly useful
10 for parenteral administration, i.e., subcutaneously, intramuscularly or intravenously. The compositions for parenteral administration will commonly comprise a solution of the immunoglobulin or a cocktail thereof dissolved in an acceptable carrier, preferably an aqueous carrier. A variety
15 of aqueous carriers can be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine, 5% glucose, human albumin solution and the like. These solutions are sterile and generally free of particulate matter. These compositions may be sterilized by conventional, well-known sterilization techniques. The
20 compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH adjusting and buffering agents, tonicity agents, toxicity adjusting agents and the like, for example sodium acetate, sodium chloride, potassium chloride, calcium
25 chloride, sodium lactate, sodium citrate, etc. The concentration of immunoglobulin in these formulations can vary widely, i.e., from the less than about 0.5%, usually at least about 1% to as much as 15 or 20% by weight and will be selected primarily based on fluid volumes, viscosities, etc., in
30 accordance with the particular mode of administration selected.

Thus, a typical pharmaceutical composition for injection could be made up to contain 1 ml sterile buffered water, and 1-10 mg of immunoglobulin. A typical composition for
35 intravenous infusion could be made up to contain 250 ml of sterile Ringer's solution, and 150 mg of immunoglobulin.

Actual methods for preparing parentally administrable compositions will be known or apparent to those skilled in the art and are described in more detail in, for example, Remington's Pharmaceutical Science, 15th ed., Mack Publishing Company, Easton, Pennsylvania (1980), which is incorporated herein by reference.

The immunoglobulins of this invention can be frozen or lyophilized for storage and reconstituted in a suitable carrier prior to use. This technique has been shown to be effective with conventional immune globulins and art-known lyophilization and reconstitution techniques can be employed. It will be appreciated by those skilled in the art that lyophilization and reconstitution can lead to varying degrees of immunoglobulin activity loss (e.g., with conventional immune globulins, IgM antibodies tend to have greater activity loss than IgG antibodies) and that use levels may have to be adjusted to compensate.

The compositions containing the present humanized immunoglobulins or a cocktail thereof can be administered for therapeutic or prophylactic treatments. In therapeutic application, compositions are administered to a patient already suffering from Verotoxin producing *E. coli* (VTEC) infection and Hemolytic Uremic Syndrome (HUS), or other toxic manifestations from VT2 antigens, in an amount sufficient to cure or at least partially arrest the toxic syndrome and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." In prophylactic applications, compositions are administered to patients at risk of infection in an amount sufficient to prevent or detectably inhibit such infection and/or toxic manifestation thereof due to VT2 antigens. Amounts effective for such uses depend upon the severity of the disease and the general state of the patient's own immune system, but generally range from about 0.1 to 5 mg/kg of immunoglobulin per patient dose being commonly used. It must be kept in mind that the materials of this invention may generally be employed in serious disease

states, that is life-threatening or potentially life-threatening situations. In such cases, in view of the minimization of extraneous substances and the lower probability of "foreign substance" rejections which are achieved by the present humanized immunoglobulins of this invention, it is possible and may be felt desirable by the treating physician to administer substantial excesses of these immunoglobulins.

Single or multiple administrations of the compositions can be carried out with dose levels and pattern being selected by the treating physician. In any event, the pharmaceutical formulations should provide a quantity of the immunoglobulin(s) of this invention sufficient to effectively treat the patient.

In particular embodiments, compositions comprising humanized immunoglobulin of the present invention may be used to detect VT2 antigens in Verotoxin producing *E. coli* (VTEC) infection and Hemolytic Uremic Syndrome (HUS) and/or in other infections producing VT2 or VT2V. Thus, a humanized immunoglobulin of the present invention, such as a humanized immunoglobulin that binds to the antigen determinant identified by the MuVTm1.1 antibody may be labeled and used to identify anatomic sites that contain significant concentrations of VT2 or VT2V. For example but not for limitation, one or more labeling moieties may be attached to the humanized immunoglobulin. Exemplary labeling moieties include, but are not limited to, radiopaque dyes, radiocontrast agents, fluorescent molecules, spin-labeled molecules, enzymes, or other labeling moieties of diagnostic value, particularly in radiologic or magnetic resonance imaging techniques.

Humanized immunoglobulins of the present invention can further find a wide variety of utilizes *in vitro*. By way of example, the immunoglobulins can be utilized for detection of VT2 antigens, or the like.

For diagnostic purposes, the immunoglobulins may either be labeled or unlabeled. Unlabeled immunoglobulins can be used in combination with other labeled antibodies (second antibodies) that are reactive with the humanized immunoglobulin, such as antibodies specific for human immunoglobulin constant regions. Alternatively, the immunoglobulins can be directly labeled. A wide variety of labels may be employed, such as radionuclides, fluors, enzymes, enzyme substrates, enzyme cofactors, enzyme inhibitors, ligands (particularly haptens), etc. Numerous types of immunoassays are available and are well known to those skilled in the art.

Kits can also be supplied for use with the subject immunoglobulins in the protection against or detection of a cellular activity or for the presence of a selected antigen. Thus, the subject immunoglobulin composition of the present invention may be provided, usually in a lyophilized form in a container, either alone or in conjunction with additional antibodies specific for the desired cell type. The immunoglobulins, which may be conjugated to a label or toxin, or unconjugated, are included in the kits with buffers, such as Tris, phosphate, carbonate, etc., stabilizers, preservatives, biocides, inert proteins, e.g., serum albumin, or the like, and a set of instructions for use. Generally, these materials will be present in less than about 5% wt. based on the amount of active immunoglobulin, and usually present in total amount of at least about 0.001% wt. based again on the immunoglobulin concentration. Frequently, it will be desirable to include an inert extender or excipient to dilute the active ingredients, where the excipient may be present in from about 1 to 99% wt. of the total composition. Where a second antibody capable of binding to the immunoglobulin is employed in an assay, this will usually be present in a separate vial. The second antibody is typically conjugated to a label and formulated in an analogous manner with the immunoglobulin formulations described above.

Human Antibodies

In another aspect of the invention, human antibodies that compete with mouse VTm1-1 for binding to verotoxin II or verotoxin II variant are provided.

a. Trioma Methodology

The basic approach and an exemplary cell fusion partner, SPAZ-4, for use in this approach have been described by Oestberg et al., Hybridoma 2:361-367 (1983); Oestberg, U.S. Patent No. 4,634,664; and Engleman et al., US Patent 4,634,666 (each of which is incorporated by reference in its entirety for all purposes). The antibody-producing cell lines obtained by this method are called triomas, because they are descended from three cells--two human and one mouse. Initially, a mouse myeloma line is fused with a human B-lymphocyte to obtain a non-antibody-producing xenogeneic hybrid cell, such as the SPAZ-4 cell line described by Oestberg, *supra*. The xenogeneic cell is then fused with an immunized human B-lymphocyte to obtain an antibody-producing trioma cell line. Triomas have been found to produce antibody more stably than ordinary hybridomas made from human cells.

The B-lymphocytes are obtained from the blood, spleen, lymph nodes or bone marrow of a human donor. *In vivo* immunization of a living human with verotoxin II or verotoxin II variant is usually undesirable because of the risk of initiating a harmful response. Thus, B-lymphocytes are usually immunized *in vitro* with these antigens or an antigenic fragment of either of these, or a cell bearing either of these. B-lymphocytes are typically exposed to antigen for a period of 7-14 days in a media such as RPMI-1640 (see Engleman, *supra*) supplemented with 10% human serum.

The immunized B-lymphocytes are fused to a xenogeneic hybrid cell such as SPAZ-4 by well known methods. For example, the cells are treated with 40-50% polyethylene glycol of MW 1000-4000, at about 37 degrees, for about 5-10 min. Cells are separated from the fusion mixture and propagated in

media selective for the desired hybrids (e.g., HAT or AH). Clones secreting antibodies having the required binding specificity are identified by assaying the trioma culture medium for the ability to bind to verotoxin II or verotoxin II variant. Triomas producing human antibodies having the desired specificity are subcloned by, e.g., the limiting dilution technique and grown *in vitro* in culture medium.

Although triomas are genetically stable they may not produce antibodies at very high levels. Expression levels can be increased by cloning antibody genes from the trioma into one or more expression vectors, and transforming the vector into a cell line such as the cell lines discussed, *infra*, for expression of recombinant or humanized immunoglobulins.

b. Transgenic Non-Human Mammals

Human antibodies reactive with verotoxin II and/or verotoxin II toxin can also be produced from non-human transgenic mammals having transgenes encoding at least a segment of the human immunoglobulin locus. Usually, the endogenous immunoglobulin locus of such transgenic mammals is functionally inactivated. Preferably, the segment of the human immunoglobulin locus includes unrearranged sequences of heavy and light chain components. Both inactivation of endogenous immunoglobulin genes and introduction of exogenous immunoglobulin genes can be achieved by targeted homologous recombination, or by introduction of YAC chromosomes. The transgenic mammals resulting from this process are capable of functionally rearranging the immunoglobulin component sequences, and expressing a repertoire of antibodies of various isotypes encoded by human immunoglobulin genes, without expressing endogenous immunoglobulin genes. The production and properties of mammals having these properties are described in detail by, e.g., Lonberg et al., WO93/12227 (1993); Kucherlapati, WO 91/10741 (1991) (each of which is incorporated by reference in its entirety for all purposes). Transgenic mice are particularly suitable. Antibodies are

obtained by immunizing a transgenic nonhuman mammal, such as described by Lonberg or Kucherlapati, *supra*. Monoclonal antibodies are prepared by, e.g., fusing B-cells from such mammals to suitable myeloma cell lines using conventional
5 Kohler-Milstein technology.

The following examples are offered by way of illustration, not by limitation. It will be understood that although the examples pertain to the HuVTm1.1 antibody, producing humanized antibodies with high binding affinity for
10 the B subunit of the VT2 antigen it is also contemplated using CDR's from other monoclonal antibodies that bind to an epitope of VT2.

EXPERIMENTAL

Example 1: Immunization of mice with VT2 toxoid

15

VT2 was prepared as described by Oku et al., Microb. Pathog., 1989, 6(2), 113-122. The VT2 toxoid was prepared by treating 1 mg of purified VT2 for 7 days at 37°C with 0.4% formaldehyde in 0.1 M phosphate buffer, pH 7.6.

20 Balb/c mice (Nippon Charles River) were immunized with VT2 toxoid (0.5µg) combined with Freund's Complete Adjuvant (Gibco BRL) by intraperitoneal (i.p.) injection. After about 4 weeks, the mice received VT2 toxoid (1 µg) combined with Freund's Complete Adjuvant by i.p. injection. Then the mice
25 received VT2 toxoid (1 µg twice, 4 µg twice, 5µg once) combined with Freund's Incomplete Adjuvant by i.p. injection sequentially at an interval of 1 to 5 weeks.

Mice were bled by cutting the tail vein and serum was collected by incubation of the blood for 30 min. at 37°C and
30 centrifugation at 3000 rpm for 10 min. The serum titer against VT2 was measured by ELISA as follows:

Each well of 96 well flat bottomed plate (Falcon 3912, Becton Dickinson) was coated with 50 µl of 1 µg/ml VT2 diluted with 0.01 M phosphate buffered saline (PBS) and
35 incubated at r.t. for 1 hr.

Each well was washed 3 times with PBS - 0.05% Tween 20.

Each well was blocked with PBS - 3% BSA at r.t. for 1 hr.
Each well was washed 3 times with PBS - 0.05% Tween 20.
50µl of serum serially diluted with PBS was added to each well and incubated at r.t. for 1 hr.

5 Each well was washed 3 times with PBS - 0.05% Tween 20.
To each well was added 50µl of alkaline phosphatase conjugated goat anti-mouse IgG (ZYMED) diluted ($\times 1000$) with PBS - 3% BSA and incubated at r.t. for 1 hr.
Each well was washed 3 times with PBS - 0.05% Tween 20.

10 To each well was added p-nitrophenyl phosphate (PNPP:Wako Chemicals) and incubated at r.t. for 1 hr.
The absorption at 405 nm was measured and recorded for each well.

15 Example 2. Construction of hybridoma by cell fusion method

Mice whose serum contained antibodies reactive against VT2 were chosen and splenectomized. 5×10^7 spleen cells and 5×10^6 P3 X 63 Ag8U.1 (P3U1) mouse myeloma cells were
20 combined and washed once with RPMI 1640 medium and centrifuged at 1500 rpm for 5 min. The cell pellet was gently dispersed and resuspended in 1 ml of polyethyleneglycol (PEG) solution (containing RPMI 1640 medium 5.75 ml + PEG 3.5 ml + dimethylsulfoxide 0.75 ml) and gently rotated for 2 min. Then
25 1 ml of RPMI medium was added and rotated for 2 min. After that, 2 ml of RPMI medium was added and rotated for 2 min. 4 ml of GIT-HAT medium (95µM hypoxanthine, 0.4 µM aminopterin, 1.6 µM thymidine, 5% FCS) was added and rotated for 2 min. Then 8 ml of GIT-HAT medium was added and rotated for 2 min.
30 After 30 min. incubation at 37°C the cell suspension was delivered into each well of the 96 well flat bottomed plate inoculated with 10^4 mouse peritoneal macrophage/well. The plate was incubated at 37°C in 5% CO₂ - 95% air incubator for one week. Then half of the medium in each well was replaced
35 with fresh GIT-HT medium (GIT-HAT medium without aminoferin)

and the plate was incubated for about one week to grow hybridoma cells.

Example 3: Screening of mouse hybridoma cells secreting antibody against VT2

Screening was done by selecting hybridoma cells secreting antibodies which bind to VT2 and neutralize VT2. Hybridoma supernatant was used for the screening. Binding activity was measured according to the ELISA method described in Example 1. VT2 neutralization activity was measured according to the following method.

To each well of a 96 well plate was added 30 μ l of hybridoma supernatant and 30 μ l of 200 pg/ml VT2 in 10% FCS-MEM.

The plate was incubated at 37°C for 1 hr.

50 μ l of the above solution was added to each well of a 96 well flat bottomed plate on which a Vero cell (ATCC) was inoculated (4×10^4 cells/well).

The plate was incubated at 37°C for 4 days in a 5% CO₂ - 95% air incubator.

100 μ l of a 0.014% neutral red solution was added to each well.

The plate was incubated at 37°C for 1 hr. to stain live cells.

After washing the well with PBS, 100 μ l of 50% ethanol solution containing 1% acetic acid (ETOH-AcOH) solution was added to each well.

The absorption at 550 nm was measured and recorded for each well.

Example 4: Cloning of hybridoma cells

Cloning of hybridoma cells was conducted by limiting dilution. One to two hybridoma cells were added to each well of a 96 well flat bottomed plate previously inoculated with 10^4 mouse peritoneal macrophages as the feeder cells. After about 2 weeks, the supernatant was measured for the binding to

VT2 and neutralization against VT2 as described in Examples 1 and 3, respectively. By repeating this cloning procedure several times as described above, a hybridoma cell clone secreting a monoclonal antibody (MuVTm1.1) which neutralizes VT2 was established.

Example 5: Purification of MuVTm1.1

The hybridoma cells secreting MuVTm1.1 were adapted to eRDF basal serum free medium containing insulin, transferrin, ethanolamine, and selenite. The culture supernatant was passed over a Protein G sepharose column (Pharmacia) and the antibody eluted using standard methods. The purity of the antibody was verified by polyacrylamide gel electrophoresis (PAGE), and its concentration was determined by SRID. The purified MuVTm1.1 was tested for its ability to neutralize VT2 variants as described in Example 3. The results are presented in the table below and in Figure 7.

TABLE 1

Neutralizing Activity of MuVTm1.1 Against Various VT2 Variants

Strains	Origin	Toxin Type	Neutralization
<i>E. coli</i> 0157 V50	Human	VT2vh	+
<i>E. coli</i> 0157 V354	Human	VT2vh	+
<i>E. coli</i> 0157 V601	Human	?	+
<i>E. coli</i> 0157:H7	Human	VT2, VT2vx1*	+
TK40			
<i>E. coli</i> 0157:H7	Human	VT2vx1	+
TK51			
<i>E. coli</i> 091:H21	Human	VT2vha, VT2vhb	+

* VT2vx is a novel subtype of VT2 variant

Example 6: Cloning and sequencing of mouse VTm1.1
variable region cDNAs

Mouse VTm1.1 (MuVTm1.1) heavy and light chain variable
region cDNAs were cloned from mRNA isolated from hybridoma
5 cells using anchored PCR (Co et al., J. Immunol. 148: 1149
(1992)). The 5' primers used annealed to the poly-dG tails
added to the cDNA, and the 3' primers to the constant
regions. The amplified gene fragments were then inserted
into the plasmid pUC18. Nucleotide sequences were
10 determined from several independent clones for both V_L and
 V_H cDNA. For the heavy chain, a single, unique sequence was
identified, typical of a mouse heavy chain variable region.
For the light chain, two unique sequences, both homologous
to mouse light chain variable region sequences, were
15 identified. However, one sequence was not functional
because of a missing nucleotide that caused a frame shift at
the V-J junction, and was identified as the non-productive
allele. The other sequence was typical of a functional
mouse kappa chain variable region. The variable region cDNA
20 sequences of the heavy chain and the functional light chain
and the translated amino acid sequences are shown in Figure
1. The mouse V_L sequence belongs to Kabat's mouse kappa
chain subgroup V. The mouse V_H belongs to Kabat's heavy
chain subgroup III(D).

Example 7: Design of humanized VTm1.1 variable regions

To retain the binding affinity of the mouse antibody in
the humanized antibody, the general procedures of Queen et
al. were followed (Queen et al. Proc. Natl. Acad. Sci. USA
30 86: 10029 (1989) and U.S. Patent Nos. 5,585,089 and
5,693,762). The choice of framework residues can be
critical in retaining high binding affinity. In principle,
a framework sequence from any human antibody can serve as
the template for CDR grafting; however, it has been
35 demonstrated that straight CDR replacement into such a

framework can lead to significant loss of binding affinity to the antigen (Glaser et al., J. Immunol. 149:2606 (1992); Tempest et al., Biotechnology 9:266 (1992); Shalaby et al., J. Exp. Med. 17:217 (1992)). The more homologous a human antibody is to the original mouse antibody, the less likely will the human framework introduce distortions into the mouse CDRs that could reduce affinity. Based on a sequence homology search against the Kabat database (Kabat et al., Sequences of Proteins of Immunological Interest, 5th ed., U.S. Department of Health and Human Services, 1991), the human antibody GF4 was chosen as providing good framework homology to the MuVTm1.1 antibody. Other highly homologous human antibody chains would also be suitable to provide the humanized antibody framework, especially kappa light chains from human subgroup III and heavy chains from human subgroup III as defined by Kabat.

The computer programs ABMOD and ENCAD (Levitt et al., J. Mol. Biol. 168: 595 (1983)) were used to construct a molecular model of the VTm1.1 variable domain, which was used to locate the amino acids in the VTm1.1 framework that are close enough to the CDRs to potentially interact with them. To design the humanized VTm1.1 heavy and light chain variable regions, the CDRs from the mouse VTm1.1 antibody were grafted into the framework regions of the human GF4 antibody. At framework positions where the computer model suggested significant contact with the CDRs, the amino acids from the mouse antibody were substituted for the original human framework amino acids. For humanized VTm1.1, this was done at residues 29, 30, 49 and 98 of the heavy chain and at residue 49 of the light chain. Furthermore, framework residues that occurred only rarely at their positions in the database of human antibodies were replaced by a human consensus amino acid at those positions. For humanized VTm1.1 this was done at residues 1, 4, 5, 79, 89 and 93 of the heavy chain and at residues 3, 4, 19, 76, 79 and 85 of the light chain.

The sequence of the humanized VTm1.1 antibody heavy chain and light chain variable regions, including the initiation codon and the signal peptide sequences, are shown in Figure 2. However, many of the potential CDR-contact residues are amenable to substitutions of other amino acids that may still allow the antibody to retain substantial affinity to the antigen. The following table lists a number of positions in the framework where alternative amino acids may be suitable (LC = light chain, HC = heavy chain).

TABLE 2

Position	Humanized VTm1.1	Alternatives
LC-49	K	Y
HC-29	F	S
HC-30	S	K
HC-49	A	S
HC-98	R	K

Likewise, many of the framework residues not in contact with the CDRs in the humanized VTm1.1 heavy and light chains can accommodate substitutions of amino acids from the corresponding positions of the human GF4 antibody, from other human antibodies, from the mouse VTm1.1 antibody, or from other mouse antibodies, without significant loss of the affinity or non-immunogenicity of the humanized antibody. The following table lists a number of additional positions in the framework where alternative amino acids may be suitable.

TABLE 3

Position	Humanized VTm1.1	Alternatives
LC-3	V	L
LC-4	L	M
LC-19	A	V
LC-76	S	N
LC-79	E	Q
LC-85	V	L, M
HC-1	E	Q
HC-4	L	V
HC-5	V	L
HC-79	L	V
HC-89	E	D
HC-93	V	M, I

Selection of various alternative amino acids may be used to produce versions of humanized VTm1.1 that have varying combinations of affinity, specificity, non-immunogenicity, ease of manufacture, and other desirable properties. Thus, the examples in the above tables are offered by way of illustration, not of limitation.

Example 8: Construction of humanized VTm1.1

Once the humanized variable region amino acid sequences had been designed as described above, genes were constructed to encode them, including signal peptides, splice donor signals and appropriate restriction sites (Figure 2). The light and heavy chain variable region genes were constructed and amplified using eight overlapping synthetic oligonucleotides ranging in length from approximately 65 to 80 bases, as shown in Figure 3 (see He et al. J. Immunol. 160: 1029 (1998)). The oligos were annealed pairwise and extended with the Klenow fragment of DNA polymerase I, yielding four double-stranded fragments. The resulting fragments were denatured, annealed, and extended with Klenow, yielding two fragments. These fragments were denatured, annealed pairwise, and extended once again,

yielding a full-length gene. The resulting product was amplified by polymerase chain reaction (PCR) using Taq polymerase, gel-purified, digested with XbaI, gel-purified again, and subcloned into the XbaI site of the pVk or pVg1 expression vector. The pVk and pVg1 vectors for the
5 respective light chain and heavy chain expression have been previously described (see Co et al., J. Immunol. 148:1149 (1992)).

The structure of the final plasmids were verified by
10 nucleotide sequencing and restriction mapping. All DNA manipulations were performed by standard methods well-known to those skilled in the art.

To construct a cell line producing humanized VTm1.1, the heavy chain and light chain plasmids were transfected
15 into the mouse myeloma cell line Sp2/0-Ag14 (ATCC CRL 1581). Before transfection, the heavy and light chain-containing plasmids were linearized using FspI. Approximately 20 μ g of each plasmid was transfected into 1×10^7 cells in PBS. Transfection was by electroporation using a Gene Pulser
20 apparatus (BioRad) at 360 V and 25 μ FD capacitance according to the manufacturer's instructions. The cells from each transfection were plated in four 96-well tissue culture plates, and after two days, selection medium (DMEM, 10% FCS, 1 x HT supplement (Sigma), 0.25 mg/ml xanthine, 1 μ g/ml
25 mycophenolic acid) was applied.

After approximately two weeks, the clones that appeared were screened for antibody production by ELISA. Antibody from a high-producing clone was prepared by growing the cells to confluency in regular medium (DMEM with 10% FCS),
30 then replacing the medium with a serum-free medium (Hybridoma SMF; Gibco) and culturing until maximum antibody titers were achieved in the culture. The culture supernatant was run through a protein A-Sepharose column (Pharmacia); antibody was eluted with 0.1 M Glycine, 100 mM
35 NaCl, pH 3, neutralized and subsequently exchanged into phosphate-buffered saline (PBS). The purity of the antibody

was verified by analyzing it on an acrylamide gel, and its concentration was determined by an OD₂₈₀ reading, assuming 1.0 mg of antibody protein has an OD₂₈₀ reading of 1.4.

5 Example 9: Properties of humanized VTm1.1

Affinity measurement

10 The affinity of MuVTm1.1 and HuVTm1.1 antibody for verotoxin II (VT2) was determined by competitive binding with biotinylated MuVTm1.1 antibody. The procedure for the experiment is described below:

15 Coated Dynatech Laboratories Immulon 2 96-well plate (part #0110103455) with 50 μ l VT2 solution (0.2 μ g/ml in PBS) per well. Incubated at 37 °C for 2 hr with gentle shaking.

Aspirated off the VT2 solution. Washed each well 4 times with 400 μ l of wash buffer (0.1% Tween 20 in PBS).

20 Blocked each well with 400 μ l of Pierce SuperBlock Blocking Buffer in PBS (cat #37515) at room temperature for 30 min.

Aspirated off the blocking solution. Washed each well 4 times with 400 μ l of wash buffer.

25 In each well added 20 ng biotinylated MuVTm1.1 antibody mixed with various concentrations of unlabeled mouse or Hu VTm1.1 antibodies in a total volume of 100 μ l in binding buffer (1% BSA, 0.1% Tween 20 in PBS). Incubated at 4 °C overnight with gentle shaking. Aspirated off the antibody solutions.
30 Washed each well 4 times with 400 μ l of wash buffer. Added 100 μ l of peroxidase-conjugated streptavidin (Biosource cat #5NN2004) (1:500 dilution in binding

buffer) to each well. Incubated at 37 °C for 1 hr with gentle shaking.

Aspirated off the streptavidin solution. Washed each well 6 times with 400 μ l of wash buffer.

Added 100 μ l /well peroxidase substrate (BioRad #172-1064). Incubated at room temperature until the color developed. Read the plate in Molecular Devices ELISA plate reader at 415 nm.

The result, shown in Figure 4, demonstrated that the humanized VTml.1 antibody competes well against the biotinylated mouse antibody in the competitive ELISA assay, within a factor of 2 when compared to the mouse antibody.

The affinity of HuVTml.1 and MuVTml.1 is also calculated using the BIAcore method. The calculated K_D for the HuVTml.1 is approximately 3.6×10^{-9} M versus 1.9×10^{-9} M for the Mu VTml.1 as shown in the Table below:

TABLE 4

	MuVTml.1	HuVTml.1
K_a	$9.9 \times 10^4 \text{ M}^{-1} \text{ S}^{-1}$	$9.5 \times 10^4 \text{ M}^{-1} \text{ S}^{-1}$
K_d	$1.9 \times 10^{-4} \text{ S}^{-1}$	$3.4 \times 10^{-4} \text{ S}^{-1}$
K_D	$1.9 \times 10^{-9} \text{ M}$	$3.6 \times 10^{-9} \text{ M}$

Example 10: In vitro neutralizing activity of HuVTml.1

The neutralizing activity of HuVTml.1 is also measured compared to mouse MuVTml.1 in an *in vitro* assay described below:

Human kidney derived cell line (ACHN) was inoculated onto a 96-well plate at 1×10^4 cells/well and incubated at 37°C for 24 hr.

Medium was aspirated and 50 μ l of a mixed solution of 30 μ l of 540 pg/ml VT2 solution diluted with 10% FCS-MEM and 30 μ l of test antibody serially diluted with 10% FCS-MEM (mouse or humanized VTml.1) preincubated at 37°C for 1 hr

was added onto each well and incubated at 37°C for four days.

100µl of 0.028% neutral red in medium was added to each well and incubated at 37°C for 1 hr (Mullbacher, A. et al., 1984, Journal of Immunological Methods, 68, 205-215).

Solution was aspirated. Each well was washed twice with 150 µl of PBS.

100µl of 50% EtOH/1% AcOH was added to each well and incubated at room temperature for 5-10 min.

The absorption value at 550 nm was recorded for each well using a Molecular Devices ELISA plate reader.

Neutralizing activity was calculated according to the following formula:

$$\text{Neutralization (\%)} = \frac{\text{OD(VT2 + Ab)} - \text{OD (VT2 alone)}}{\text{OD (medium alone)} - \text{OD (VT2 alone)}} \times 100$$

The neutralizing activity of HuVTm1.1 compared to MuVTm1.1 is shown in Figures 5. The ED50 for the HuVTm1.1 is 0.33 µg/ml versus 0.2 µg/ml for MuVTm1.1. The neutralization activity of HuVTm1.1 to VT2 variants is shown below:

TABLE 5

In vitro VT2variants neutralizing assay with HuVTm1.1

Strains	Origin	Toxin Type	ED50 (µg/ml)
<i>E. coli</i> 0157	Human	VT2	0.33
<i>E. coli</i> 0157 (V50)	Human	VT2vh	0.36
<i>E. coli</i> 0157 (V354)	Human	VT2vh	0.39
<i>E. coli</i> 0157 (V601)	Human	VT2v	0.32
<i>E. coli</i> 0157:H7 (TK 40)	Human	VT2, VT2vx*	0.35

* VT2vx: Novel subtype of VT2 variant

Example 11: Analysis of VT2 antigen recognized

VT2 was reduced, the A and B subunits were separated by polyacrylamide gel containing SDS and transferred to a

nitrocellulose membrane (Bio Rad) by Western Blotting method. The nitrocellulose membrane was blocked overnight in 3% BSA in PBS and then reacted with 10µg/ml of HuVTm1.1 or rabbit anti-VT2 serum diluted with 3% BSA in PBS for 1 hr. at r.t. The membrane was washed 6 times with 1% BSA-PBS and reacted with 25µg/ml alkaline phosphatase conjugated goat anti-human IgG (TAGO) or alkaline phosphatase conjugated goat anti-rabbit IgG (TAGO) diluted with 3% BSA in PBS for 1 hr. at r.t. The membrane was washed 6 times with 1% BSA in PBS and reacted with NBT (Nitro-Blue Tetrazolium Chloride) - BCIP (5-Bromo-4-Chloro-3'-Indolyephosphate p-Toluidine salt) solution [PIERCE] for 10 min. at r.t. The result shown in Figure 6 indicates that HuVTm1.1 principally binds to the B subunit of VT2.

Example 12: In vivo activity of HuVTm1.1 against VT2

DdY mice (Nippon SLC) were injected via the intravenous route with 4.8 µg of sterilized HuVTm1.1, MuVTm1.1 or PBS. One hour later, mice were injected via the intraperitoneal (i.p.) route with 46 ng of VT2 (corresponding to 10 LD₅₀). After a week, viability of mice was measured.

The following table shows the result. HuVTm1.1 completely protects mice from death caused by VT2.

TABLE 6

TREATMENT	SURVIVORS
PBS	100% (5/5)
VT2	0% (0/5)
HuVTm1.1 + VT2	100% (5/5)
MuVTm1.1 + VT2	100% (5/5)

From the foregoing, it will be appreciated that the humanized immunoglobulins of the present invention offer numerous advantages over other anti VT specific antibodies. In comparison to mouse monoclonal antibodies, the present humanized immunoglobulins contain substantially less non-human and potentially immunogenic amino acid sequences. This reduced likelihood of antigenicity after injection into

a human patient represents a significant therapeutic improvement.

All publications and patent applications cited above are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference. Although the present invention has been described in some detail by way of illustration and example for purposes of clarity and understanding, it will be apparent that certain changes and modifications may be practiced within the scope of the appended claims.